

# Restriction Digest Protocol

## Materials

- 10X NEBuffer (standard or unique)
- 6X Purple Load Dye (may be supplied)
- Additives if needed:
  - Adenosine 5'-Triphosphate (ATP) (supplied when required)
  - rAlbumin (supplied when required)
  - DTT (supplied when required)
  - PaqCI Activator (supplied when required)

## Materials required but not supplied:

- DNA
- Nuclease-free Water ([NEB #B1500](#))

## Overview

Restriction enzyme digests require proper quantities of DNA, enzyme, and buffer, in an appropriate reaction volume ratio. For molecular cloning, the “typical” reaction conditions below use 5-10x over digestion to compensate for variabilities in DNA sample quality and quantity. [Time-Saver™ qualified restriction enzymes](#) and [High-Fidelity \(HF®\) restriction enzymes](#) perform under a broader range of recommended conditions. Critically, individual restriction endonuclease recommended reaction times and temperatures can vary significantly.

[NEBcloner®](#) tool is strongly recommended to design reactions for successful digests.



Additional information to optimize restriction enzyme digests can be found in our usage guidelines: [Optimizing Restriction Endonuclease Reactions](#) and [Restriction Enzyme Tips](#).

## Reaction set-up:

Set up the following reaction as follows on ice.

(Please note that enzymes should always be added last.)

COMPONENTS	50 $\mu$ l REACTION	FINAL CONC. OR AMOUNT
DNA	1 $\mu$ g	
10X NEBuffer**	5 $\mu$ l	1X
Restriction Enzyme	1 $\mu$ l	
Nuclease-free Water	to 50 $\mu$ l	Varies*

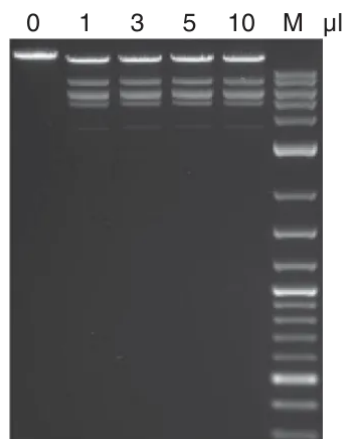
\*We recommend 5-10 units of enzyme per  $\mu$ g of DNA. Adjust according to protocol at [NEBcloner.neb.com](http://NEBcloner.neb.com). For convenience, 1  $\mu$ l can be used.

\*\*Additive (such as DTT, Activator or ATP) may be required for certain enzymes.

### Digest protocol:

1. Gently mix the reaction by pipetting up and down and microfuge briefly.
2. Incubate at temperature and time ranges recommended for the specific restriction endonuclease.
3. Stop the reaction.
  - If no further manipulation of DNA is required, use 10  $\mu$ l per 50  $\mu$ l reaction of stop solution containing EDTA, (e.g. [Gel Loading Dye, Purple \(6X\)](#), [Gel Loading Dye, Purple \(6X\)](#), no SDS, or [Gel Loading Dye, Orange \(6X\)](#))

### Gel with restriction enzyme digested Lambda DNA



50  $\mu$ l digestion reactions were set up using a range of amounts of *EcoRI-HF*<sup>®</sup> restriction enzyme, 1  $\mu$ g of Lambda DNA, and the recommended reaction buffer, resulting in no star activity in overnight digests at 37°C, even when used at higher concentrations. Marker M is the 1 kb DNA Ladder ([NEB# N3232](#)). The Monarch Spin DNA Gel Extraction Kit ([NEB #T1120](#)) is recommended for DNA gel extraction and purification

## General Guidelines

1. If further manipulation of DNA is required, use either heat inactivation at the appropriate temperature (65°C or 80°C) for 20 minutes (please note not all enzymes are inactivated by heat), phenol/chloroform extraction, or spin column purification (e.g. [Monarch<sup>®</sup> Kits for DNA Cleanup](#)) to remove the enzyme.
2. For highly concentrated DNA for downstream applications, use the Monarch Spin DNA Gel Extraction Kit ([NEB #T1120](#)) to attain high-quality DNA purification with low-elution volumes.

## Related Resources

- [Optimizing Restriction Endonuclease Reactions](#)
- [Restriction Enzyme Tips](#)
- [NEBcloner<sup>®</sup>](#)
- [NEBcutter<sup>®</sup>](#)
- [NEBioCalculator<sup>®</sup>](#)
- [REBASE<sup>®</sup>](#)
- [Double Digest Finder](#)
- [Enzyme Finder](#)
- [Competitor Cross-Reference Tools](#)