

# LunaScript RT Master Mix Kit Protocols (NEB #E3025)

## Overview

1. Mix components by vortexing briefly and spin down if necessary.
2. Prepare cDNA synthesis reaction as described below:

COMPONENT	20 $\mu$ l REACTION	FINAL CONCENTRATION
LunaScript RT Master Mix (Primer-free) (5X)	4 $\mu$ l	1X
Random Primer Mix (60 $\mu$ M) <sup>1</sup> Or d(T) <sub>23</sub> VN (50 $\mu$ M) <sup>2</sup> Or Gene-specific primer <sup>3</sup>	2 $\mu$ l 2 $\mu$ l x $\mu$ l	6 $\mu$ M 5 $\mu$ M 0.1–1 $\mu$ M (typically 0.5 $\mu$ M final)
RNA Sample <sup>4</sup>	variable	(up to 1 $\mu$ g)
Nuclease-free Water	to 20 $\mu$ l	

For no-RT control reactions, mix the following components:

COMPONENT	20 $\mu$ l REACTION	FINAL CONCENTRATION
No-RT Control Mix (Primer-free) (5X)	4 $\mu$ l	1X
Random Primer Mix (60 $\mu$ M) <sup>1</sup> Or d(T) <sub>23</sub> VN (50 $\mu$ M) <sup>2</sup> Or Gene-specific primer <sup>3</sup>	2 $\mu$ l 2 $\mu$ l x $\mu$ l	6 $\mu$ M 5 $\mu$ M 0.1–1 $\mu$ M (typically 0.5 $\mu$ M final)
RNA Sample <sup>4</sup>	variable	(up to 1 $\mu$ g)
Nuclease-free Water	to 20 $\mu$ l	

For no template controls, mix the following components:

COMPONENT	20 $\mu$ l REACTION	FINAL CONCENTRATION
LunaScript RT Master Mix (Primer-free) (5X)	4 $\mu$ l	1X

COMPONENT	20 $\mu$ l REACTION	FINAL CONCENTRATION
Random Primer Mix (60 $\mu$ M) <sup>1</sup> Or d(T) <sub>23</sub> VN (50 $\mu$ M) <sup>2</sup> Or Gene-specific primer <sup>3</sup>	2 $\mu$ l 2 $\mu$ l x $\mu$ l	6 $\mu$ M 5 $\mu$ M 0.1–1 $\mu$ M (typically 0.5 $\mu$ M final)
Nuclease-free Water	to 20 $\mu$ l	

<sup>1</sup> Random Primer Mix ([NEB #S1330](#)) is recommended for real-time qPCR detection.

<sup>2</sup> Oligo d(T)<sub>23</sub>VN ([NEB #S1327](#)) is recommended for long or full-length cDNA synthesis.

<sup>3</sup> Gene-specific primers can be used for target-specific cDNA synthesis. The final concentration is typically 0.5  $\mu$ M and can be optimized in the range of 0.1–1  $\mu$ M.

<sup>4</sup> Up to 1  $\mu$ g total RNA, 1  $\mu$ g mRNA or 100 ng specific RNA can be used in a 20  $\mu$ l reaction. To accommodate larger amounts of input RNA (> 1  $\mu$ g), the reaction should be scaled up to ensure optimum cDNA synthesis.

Incubate reactions in a thermocycler with the following steps:

PRIMERS FOR cDNA SYNTHESIS	CYCLE STEP	TEMP	TIME	CYCLES
Random Primers (e.g., Random Primer Mix)	Primer Annealing	25°C	2 minutes	1
	cDNA Synthesis	55°C	10 minutes	
	Heat Inactivation	95°C	1 minute	
Oligo-dT primers or a gene-specific primer	cDNA Synthesis	55°C	10 minutes	1
	Heat Inactivation	95°C	1 minute	

The cDNA product should be stored at -20°C. In general, the volume of cDNA product should not exceed 1/10 of the qPCR or PCR reaction volume. Where needed, up to 20% qPCR volume can be undiluted cDNA product.

For qPCR applications, we recommend using Luna Universal qPCR Master Mix ([NEB #M3003](#)) for dye-based qPCR detection and Luna Universal Probe qPCR Master Mix ([NEB #M3004](#)) for probe-based detection.

For downstream PCR, we recommend OneTaq 2X Master Mix ([NEB #M0482](#) or [NEB #M0485](#)) for PCR detection up to 5kb, Q5<sup>®</sup> Hot Start High-Fidelity 2X Master Mix ([NEB #M0494](#)) for highest fidelity, and LongAmp Taq 2X Master Mix ([NEB #M0287](#)) for high yields from longer products.