240 County Road Ipswich, MA 01938-2723 Tel 978-927-5054 Fax 978-921-1350 www.neb.com info@neb.com

New England Biolabs Certificate of Analysis

Product Name: EpiMark® Hot Start Taq DNA Polymerase

Catalog #: M0490S/L
Concentration: 5,000 units/ml

Unit Definition: One unit is defined as the amount of enzyme that will incorporate 15 nmol dNTP into acid insoluble material in 30 minutes at

75°C.

 Lot #:
 0111612

 Assay Date:
 12/2016

 Expiration Date:
 12/2018

 Storage Temp:
 -20°C

Storage Conditions: 10 mM Tris-HCl, 100 mM KCl, 1 mM DTT, 0.1 mM EDTA, 0.5 % Tween® 20, 0.5 % IGEPAL® CA-630, 50 %

Glycerol, (pH 7.4 @ 25°C)

Specification Version: PS-M0490S/L v1.0

Effective Date: 14 Jun 2016

Assay Name/Specification (minimum release criteria)	Lot #0111612
Endonuclease Activity (Nicking) - A 50 μ l reaction in ThermoPol® Reaction Buffer containing 1 μ g of supercoiled PhiX174 DNA and a minimum of 20 units of Taq DNA Polymerase incubated for 4 hours at either 37°C or 75°C results in <10% conversion to the nicked form as determined by agarose gel electrophoresis.	Pass
Inhibition of Primer Extension (Hot Start, Radioactivity Incorporation) - A 50 μl primer extension assay in ThermoPol® Reaction Buffer in the presence of 200 μM dNTPs including [³ H]-dTTP, containing 15 nM primed single-stranded M13mp18 with 2.5 units of EpiMark® Hot Start <i>Taq</i> DNA Polymerase incubated for 16 hours at 25°C yields >95% inhibition when compared to a non-hot start control reaction.	Pass
Non-Specific DNase Activity (16 Hour) - A 50 µl reaction in NEBuffer 2 containing 1 µg of T3 DNA in addition to a reaction containing Lambda-HindIII DNA and a minimum of 5 units of EpiMark® Hot Start <i>Taq</i> DNA Polymerase incubated for 16 hours at 37°C results in a DNA pattern free of detectable nuclease degradation as determined by agarose gel electrophoresis.	Pass
PCR Amplification (Hot Start 2 kb Lambda DNA) - A 50 μl reaction in EpiMark® Hot Start <i>Taq</i> Reaction Buffer in the presence of 200 μM dNTPs and 0.2 μM primers containing 20 pg Lambda DNA and 100 ng Human Genomic DNA with 1.25 units of EpiMark® Hot Start <i>Taq</i> DNA Polymerase for 30 cycles of PCR amplification results in an increase in yield of the 2 kb Lambda product and a decrease in non-specific genomic bands when compared to a non-hot start control reaction.	Pass









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Assay Name/Specification (minimum release criteria)	Lot #0111612
Phosphatase Activity (pNPP) - A 200 μl reaction in 1M Diethanolamine, pH 9.8, 0.5 mM MgCl ₂ containing 2.5 mM <i>p</i> -Nitrophenyl Phosphate (pNPP) and a minimum of 100 units <i>Taq</i> DNA Polymerase incubated for 4 hours at 37°C yields <0.0001 unit of alkaline phosphatase activity as determined by spectrophotometric analysis.	Pass
Protein Purity Assay (SDS-PAGE) - <i>Taq</i> DNA Polymerase is ≥ 99% pure as determined by SDS-PAGE analysis using Coomassie Blue detection.	Pass
qPCR DNA Contamination (<i>E. coli</i> Genomic) - A minimum of 5 units of EpiMark® Hot Start Taq DNA Polymerase is screened for the presence of <i>E. coli</i> genomic DNA using SYBR® Green qPCR with primers specific for the <i>E. coli</i> 16S rRNA locus. Results are quantified using a standard curve generated from purified <i>E. coli</i> genomic DNA. The measured level of <i>E. coli</i> genomic DNA contamination is ≤ 1 <i>E. coli</i> genome.	Pass
RNase Activity (Extended Digestion) - A 10 μ l reaction in NEBuffer 4 containing 40 ng of a 300 base single-stranded RNA and a minimum of 1 μ l of EpiMark® Hot Start Taq DNA Polymerase is incubated at 37°C. After incubation for 16 hours, >90% of the substrate RNA remains intact as determined by gel electrophoresis using fluorescent detection.	Pass
Single Stranded DNase Activity (Hot Start, FAM-Labeled Oligo) - A 50 μl reaction in ThermoPol® Reaction Buffer containing a 10 nM solution of a fluorescent internal labeled oligonucleotide and a minimum of 25 units of <i>Taq</i> DNA Polymerase incubated for 30 minutes at either 37°C or 75°C yields <10% degradation as determined by capillary electrophoresis.	Pass

Authorized by Denisa Gilaj 14 Jun 2016







Inspected by

Tony Spear-Alfonso

12 Dec 2016