

# Golden Gate Assembly Protocol for using NEBridge® Golden Gate Assembly Kit (BsmBI-v2) (NEB #E1602)

## Materials

- Gel Loading Dye, Purple (6X), no SDS

## Materials Required but not Supplied

### NEBridge® Golden Gate Assembly Kit (BsmBI-v2)

- User-defined inserts
- Competent cells
- Other materials for transformation

## Overview

Use this protocol for seamless cloning and assembly of multiple DNA fragments using the engineered Type IIS restriction enzyme BsmBI-v2 optimized for Golden Gate Assembly. It can be used for directed assembly of multiple inserts/modules and single insert/library generation cloning with single insert(s) using the Golden Gate approach.

Negative controls are not routinely done for assembly reactions but are described here for first-time users. This protocol uses the destination plasmid, pGGaselect, but others may be selected using the DNA Sequences and Maps Tool, which contains sequence files, vector maps, and cut sites.

## Protocol

1. Set up assembly reactions as follows:

REAGENT	ASSEMBLY REACTION
pGGaselect Destination Plasmid*, 75 ng/ µl	1 µl
Inserts (user provided): - if precloned** - if in amplicon form***	75 ng each plasmid 2:1 molar ratio**** (insert:vector backbone; pGGaselect = 2,155 bp; 75 ng = 0.05 pmol)
T4 DNA Ligase Buffer (10X)	2 µl
NEBridge Golden Gate Enzyme Mix (BsmBI-v2)	1–2 µl *****
Nuclease-free Water	to 20 µl*****

\* Or user provided. Consider using the [DNA Sequences and Maps Tool](#) to select destination plasmids

\*\* Precloned inserts must possess *BsmBI* restriction sites at both ends of the insert sequence and in the proper orientation.

\*\*\* Amplicon inserts must possess 5' flanking bases (6 recommended) and *BsmBI* restriction sites at both ends of the amplicon and in the proper orientation.

\*\*\*\* The [NEBioCalculator](#)<sup>®</sup> Tool can be used for molar calculations.

\*\*\*\*\* For assemblies ≤ 10 inserts, use 1 µl; for assemblies > 10 inserts, use 2 µl.

\*\*\*\*\* Can be increased to 25 µl volume if required due to DNA component volumes; add additional 0.5 µl T4 DNA Ligase Buffer (10X)

2. Choose the appropriate assembly protocol:

INSERT NUMBER	SUGGESTED ASSEMBLY PROTOCOL
For 1 insert	42°C, 5 min (cloning) or 42°C, 1 hr (library preparation) → 60°C, 5 min
For 2–10 inserts	(42°C, 1 min → 16°C, 1 min) x 30 → 60°C, 5 min
For 11–20+ Inserts	(42°C, 5 min → 16°C, 5 min) x 30 → 60°C, 5 min

To learn more about performing complex assemblies, visit [NEBridge Golden Gate Assembly](#).

## Resources:

- [Learn more about NEBridge for Golden Gate Assembly](#)
- [NEBridge Golden Gate Assembly Tool](#)
- [DNA Sequences and Maps Tool](#)
- [NEBioCalculator Tool](#)
- [NEBridge Ligase Fidelity Tools](#)
- [Applications of Ligase Fidelity Data & Tools](#)
- [Breaking through the Limitations of Golden Gate Assembly](#)