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## New England Biolabs Product Specification

Product Name: Salt-T4TM DNA Ligase

Catalog #: M0467S/L

Concentration: 400,000 units/ml

Unit Definition: One unit is defined as the amount of enzyme required to give 50% ligation of 6 µg of Lambda-HindIII DNA in 30 minutes

at 25°C in a total reaction volume of 20 µl in 1X T4 DNA Ligase Reaction Buffer supplemented with 100 mM NaCl.

Shelf Life: 24 months
Storage Temp: -20°C

Storage Conditions: 10 mM Tris-HCl, 50 mM KCl, 1 mM DTT, 0.1 mM EDTA, 50 % Glycerol, (pH 7.4 @ 25°C)

Specification Version: PS-M0467S/L v1.0

Effective Date: 29 Oct 2019

## Assay Name/Specification (minimum release criteria)

DNase Activity (Labeled Oligo, 3' extension) - A 50 µl reaction in CutSmart® Buffer containing a 20 nM solution of a fluorescent labeled double-stranded oligonucleotide containing a 3' extension and a minimum of 2000 units of Salt-T4<sup>TM</sup> DNA Ligase incubated for 16 hours at 37°C yields <5% degradation as determined by capillary electrophoresis.

DNase Activity (Labeled Oligo, 5' extension) - A 50 μl reaction in CutSmart® Buffer containing a 20 nM solution of a fluorescent labeled double-stranded oligonucleotide containing a 5' extension and a minimum of 2000 units of Salt-T4<sup>TM</sup> DNA Ligase incubated for 16 hours at 37°C yields <5% degradation as determined by capillary electrophoresis.

Double Stranded DNase Activity (Labeled Oligo) - A 50 μl reaction in CutSmart® Buffer containing a 20 nM solution of a fluorescent labeled double-stranded oligonucleotide containing a blunt end and a minimum of 2000 units of Salt-T4<sup>TM</sup> DNA Ligase incubated for 16 hours at 37°C yields <5% degradation as determined by capillary electrophoresis.

Endonuclease Activity (Nicking) - A 50  $\mu$ l reaction in NEBuffer 1 containing 1  $\mu$ g of supercoiled PhiX174 DNA and a minimum of 400 units of Salt-T4<sup>TM</sup> DNA Ligase incubated for 4 hours at 37°C results in <10% conversion to the nicked form as determined by agarose gel electrophoresis.

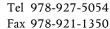
Non-Specific DNase Activity (16 Hour) - A 50  $\mu$ l reaction in NEBuffer 1 containing 1  $\mu$ g of CIP-treated Lambda-HindIII DNA and a minimum of 400 units of Salt-T4<sup>TM</sup> DNA Ligase incubated for 16 hours at 37°C results in a DNA pattern free of detectable nuclease degradation as determined by agarose gel electrophoresis.

Protein Concentration (A280) - The concentration of Salt-T4<sup>TM</sup> DNA Ligase is 0.4 mg/ml +/- 10% as determined by UV absorption at 280 nm. Protein concentration is determined by the Pace method using the extinction coefficient of 57,675 and molecular weight of 56,894 daltons for Salt-T4<sup>TM</sup> DNA Ligase (Pace, C.N. et al. (1995) Protein Sci., 4, 2411-2423).









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Assay Name/Specification (minimum release criteria)

Protein Purity Assay (SDS-PAGE) - Salt-T4<sup>TM</sup> DNA Ligase is  $\geq$  95% pure as determined by SDS-PAGE analysis using Coomassie Blue detection.

qPCR DNA Contamination (E. coli Genomic) - A minimum of 400 units of Salt-T4<sup>TM</sup> DNA Ligase is screened for the presence of *E. coli* genomic DNA using SYBR® Green qPCR with primers specific for the *E. coli* 16S rRNA locus. Results are quantified using a standard curve generated from purified *E. coli* genomic DNA. The measured level of *E. coli* genomic DNA contamination is  $\leq 1$  *E. coli* genome.

RNase Activity (Extended Digestion) - A 10  $\mu$ l reaction in NEBuffer 4 containing 40 ng of a 300 base single-stranded RNA and a minimum of 1  $\mu$ l of Salt-T4<sup>TM</sup> DNA Ligase is incubated at 37°C. After incubation for 16 hours, >90% of the substrate RNA remains intact as determined by gel electrophoresis using fluorescent detection.

Single Stranded DNase Activity (FAM-Labeled Oligo) - A 50  $\mu$ l reaction in CutSmart® Buffer containing a 20 nM solution of a fluorescent internal labeled oligonucleotide and a minimum of 2000 units of Salt-T4<sup>TM</sup> DNA Ligase incubated for 16 hours at 37°C yields <5% degradation as determined by capillary electrophoresis.

Kuk Kotum

Date 29 Oct 2019

Derek Robinson
Director of Quality Control





