Labeling of *Escherichia coli* Expressed SNAP-tag® Fusion Proteins

**Introduction**

*Escherichia coli (E. coli)* is the leading host for expression of proteins, primarily due to its ease of use. SNAP-tag technology provides biologists with a versatile tool for expression and labeling of proteins in order to characterize their functions and interactions (1–3). New England Biolabs has introduced the pSNAP-tag (T7)-2 vector which allows a protein of interest to be fused to the N-terminus or C-terminus of an *E. coli* codon optimized SNAP-tag (Figure 1). The expression of SNAP-tagged fusion proteins are under the control of the IPTG-inducible T7/lac promoter and can be efficiently expressed using an *E. coli* T7 expression strain of your choice (4). Using this highly flexible system, a single gene construct creates a fusion protein that can be covalently labeled with a fluorophore of your choice, biotin or beads for fluorescent imaging, pull-down or other biochemical analysis (5,6).

**Materials**

- pSNAP-tag(T7)-2 Vector (#N9181)
- SNAP-Cell® TMR-Star (#S9105)
- *E. coli* T7 Express (#C2566)

**Additional Materials**

- Growth medium
- Ampicillin
- IPTG
- Phosphate Buffered Saline (PBS)
- Dithiothreitol (DTT)
- 3X SDS Sample buffer (#B7709 or #B7703)

**General Protocol**

**Cloning:** The protein of interest can be expressed with the SNAP-tag (20 kDa) as either a N- or C-terminal in-frame fusion.

1. For fusion to the C-terminus of the SNAP-tag, subclone your gene of interest into the 3´ multiple cloning site (MCS) of the pSNAP-tag(T7)-2 vector and include a stop codon in the insert at the C-terminus of the fusion gene.
2. For expression at the N-terminus of the SNAP-tag, the target gene should be cloned in-frame to the SNAP-tag and no stop codon should be included in the insert.

**Expression:** pSNAP-tag(T7)-2 can be used for expression in all NEB T7 Express strains: T7 Express Competent E. coli (NEB #C2566), T7 Express Competent E. coli (NEB #C3016), T7 Express lyS Competent E. coli (NEB #C3010), T7 Express lysY Competent E. coli (NEB #C3013) and BL21(DE3) (NEB #C2527).

1. Grow the transformed cells at 37°C until OD$_{600}$ ~0.5 (or 0.6–0.8 for 15°C induction).
2. Induce the cells with 0.4 mM IPTG at 30°C for 3–5 hours or 15°C for 16 hours.
3. Harvest the bacteria by centrifuging the culture for 15 min at 5000 x g.
4. The bacterial pellets may be stored frozen at -20°C prior to lysis and SDS-PAGE analysis of protein expression. (See Figure 2A)

**Labeling in cell lysate:**
1. Resuspend the cells in 1X PBS supplemented with 1 mM DTT and lyse the cells by sonication.
2. Sonicate to lyse the cells and add 10 μM of the substrate.
3. Incubate at 37°C for 30 minutes.
4. Add 3X SDS Sample buffer and conduct SDS-PAGE.
5. Perform in-gel fluorescent detection using a Typhoon™ 9400 Imager (Figure 2B and 2C).

**Note:** It is possible to label a SNAP-tag fusion protein at a wide range of temperatures (4°–65°C); however, optimization of various parameters (labeling time, substrate concentration etc.) should be conducted. The expressed SNAP-tag fusion protein can also be purified and subsequently labeled in 1X PBS buffer supplemented with 1 mM DTT.

**Results**

**Summary**

This note demonstrates the high level expression of SNAP-tag and three SNAP-tag fusion proteins from the pSNAP-tag (T7)-2 vector in E. coli T7 Express host cells. Specific protein labeling was readily performed in cell lysates by addition of a fluorescent substrate, either SNAP-Vista Green or SNAP-Cell TMR Star. This bacterial SNAP-tag expression system provides a versatile tool in site-specific labeling of proteins for a variety of applications.

**References:**

**Figure 2.** Expression and labeling of SNAP-tag fusions in E. coli T7 Express host cells. SNAP-tag(T7)-2 plasmids expressing SNAP-tag (23 kDa, lane 2), SNAP-tag-6XHis-tag–chitin binding domain (27 kDa, lane 3), MCP-SNAP-tag-6XHis tag (31 kDa, lane 4) and maltose binding protein-SNAP-tag (63 kDa, lane 5) were transformed into E. coli T7 Express. Induction was carried out at 30°C for 4 hours. Uninduced cells are present in lane 1. The resulting cell lysates were subjected to SDS-PAGE and stained with Coomassie blue (A), labeled with SNAP-Vista Green (B) or labeled with SNAP-Cell TMR Star (C). The fluorescent signals were detected by Typhoon Imager 9400 with the excitation/emission filter sets of 488/520 nm (B) and 532/580 nm (C). M contains the Protein Ladder (NEB #P7703).