

- If dilution of enzyme for storage is needed, we recommend using Diluent A (NEB #B8001).
- Reactions with SplintR should be performed between 16–37°C. We recommend initial testing be performed at 25°C.
- We suggest a reaction time in the range of 10–60 minutes, with 15 minutes being ideal for many applications.
- The enzyme is supplied as a 10.5 μM solution. We suggest maintaining the enzyme below 1 μM in the reaction with a suggested range of 100 nM to 1 μM . For many applications, starting with a 2-fold excess of enzyme over ligatable ends is ideal. For example, in the experiment described in the accompanying figure a substrate concentration of 100 nM was found to be ideal. In that workflow, 250 nM enzyme gave complete ligation in 15 minutes on all sequences tested.
- If the reaction is not proceeding as efficiently as desired, we strongly recommend extending the incubation time rather than increasing the concentration of enzyme in the reaction beyond 1 μM .
- An alternative option for recalcitrant substrates is to use a low concentration of ATP. A low ATP buffer can give higher yields of ligation product for substrates that have low ligation efficiencies in the standard SplintR Ligase Reaction Buffer, such as substrates with runs of G:C base pairs at the ligation junction. We suggest 1X T4 RNA Ligase Reaction Buffer (NEB #B0216) supplemented with ATP (NEB #P0756) to a final concentration of 10 μM .
- We recommend an RNA splint of at least 20 complementary bases. We have found 10 bp of dsDNA/RNA on either side of the junction to be sufficient for all substrates tested. The splint does not have to be centered on the ligation junction, however, with as few as four bases on one side of the junction giving complete ligation for a splint with 20 bases of total complementarity, depending on substrate sequence. If regions of overlap < 10 bp are desired, some testing will be required to determine the minimum length of the ds region for your specific sequence.



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