The PreCR™ Repair Mix is an enzyme cocktail that is very active at sealing nicks in DNA but does not ligate fragmented DNA. In fact, the ligase present in the mix will not repair 8-oxo-7,8-dihydro-2'deoxyadenosines or hydroxyl group. The PreCR Repair Mix will not repair all ranges of DNA damages, including those that block PCR (e.g. apurinic/apyrimidinic sites, thymine dimers, nicks and gaps) and those that are mutagenic (e.g. deaminated cytosine and 8-oxo-guanine). In addition, it will remove a variety of moieties from the 3’ end of DNA leaving a hydroxyl group. The PreCR Repair Mix will not repair damages that inhibit/interfere with PCR. For example, it will not repair 8-oxo-7,8-dihydro-2’deoxyadenosines or fragmented DNA. In fact, the ligase present in the mix is very active at sealing nicks in DNA but does not ligate blunt ends or nicks near a mismatch effectively. This limits the possibility of chimeric gene formation as a product of ligation. The PreCR Repair Mix can be used in conjunction with any thermophilic polymerase.

### Table 1: Types of DNA Damage

<table>
<thead>
<tr>
<th>DNA Damage</th>
<th>Cause</th>
<th>Can it be repaired by the PreCR Repair Mix?</th>
</tr>
</thead>
<tbody>
<tr>
<td>abasic sites</td>
<td>hydrolysis</td>
<td>yes</td>
</tr>
<tr>
<td>nicks</td>
<td>hydrolysis</td>
<td>yes</td>
</tr>
<tr>
<td>thymidine dimers</td>
<td>nucleases</td>
<td>yes</td>
</tr>
<tr>
<td>blocked 3’-ends</td>
<td>UV radiation</td>
<td>yes</td>
</tr>
<tr>
<td>oxidized guanine</td>
<td>oxidation</td>
<td>yes</td>
</tr>
<tr>
<td>oxidized pyrimidines</td>
<td>oxidation</td>
<td>yes</td>
</tr>
<tr>
<td>deaminated cytosine</td>
<td>hydrolysis</td>
<td>yes</td>
</tr>
<tr>
<td>fragmentation</td>
<td>hydrolysis/nucleases shearing</td>
<td>no</td>
</tr>
<tr>
<td>Protein-DNA crosslinks</td>
<td>formaldehyde</td>
<td>no</td>
</tr>
</tbody>
</table>

Source: Each of the recombinant proteins present in the PreCR Repair Mix has been expressed in *E. coli.*

Application:
- Repair of DNA prior to its use as a template in PCR or other DNA technologies.

Supplied in: 20 mM Tris-HCl (pH 7.5), 100 mM NaCl and 50% glycerol.

**Reagents Supplied:**
1X PreCR Repair Mix
10X ThermPol Reaction Buffer
100X NAD⁺ solution
UV damaged Lambda DNA (0.5 ng/µl)
L1 Primer Mix (20 µM)
10X Purified BSA (10 mg/ml)

**PreCR Repair Mix components:**
- Taq DNA Ligase
- Endonuclease IV
- Bst DNA Polymerase
- Fpg
- Uracil-DNA Glycosylase (UDG)
- T4 PDG (T4 Endonuclease V)
- Endonuclease VIII

**Quality Controls:**
- No

**Heat Inactivation:**
- No

**Reactions Conditions:**
- DNA template, 100 µM dNTPs, 1X ThermPol Reaction Buffer, 1X NAD⁺ and 1 µl PreCR Repair Mix in a total reaction volume of 50 µl. Incubate at 37°C.

- 1X ThermPol Reaction Buffer:
  - 10 mM KCl
  - 10 mM (NH₄)₂SO₄
  - 20 mM Tris-HCl
  - 2 mM MgSO₄
  - 0.1% Triton X-100
  - pH 8.8 @ 25°C

- 1X NAD⁺ Solution:
  - 0.5 mM NAD⁺

**Quality Controls:**
- The PreCR repair mix was tested for the ability to rescue successful amplification from UV damaged Lambda DNA (NEB #N3017). The mix was also tested for the ability to cleave oligonucleotide substrates containing thymine glycol, deoxyuracil, or 8-oxo-guanine in the absence of dNTPs. The absence of dNTPs prevents full repair and allows the presence of specific repair-targeting activities to be evaluated.

(see other side)
Standard Reaction Protocol:
In this procedure the PreCR repair and PCR reactions are performed in the same tube. This is convenient if using ThermPol buffer for the subsequent PCR reaction. Other PCR buffers may work in the repair reaction. If using a buffer other than ThermPol and optimal repair is desired, then the Sequential Reaction Protocol is recommended (see below).

The following protocol is recommended for a 50 µl reaction volume:

1. At room temperature, combine 1X ThermPol Buffer, 100 µM dNTPs, 1X NAD+, damaged template DNA and H2O to 46 µl.
2. Add 1 µl of the PreCR Repair Mix, and mix gently.
3. Incubate the repair reaction for 15–20 minutes at 37°C.
4. Place the reactions on ice.
5. Add the primers, a second aliquot of dNTPs (another 100 µM) and the PCR polymerase directly to the repair reaction mix.
6. Proceed with the PCR amplification protocol.

Notes: The extent of damage that a particular DNA template contains is variable. Therefore, it is advisable to try different amounts of template in the amplification reaction. If possible, a titration range of 200, 100, 50 and 25 ng of template DNA in the amplification reaction is recommended.

For some types or extents of damage, optimal repair occurs at 4°C and a 15 minute incubation. In certain cases we have found that a 4°C overnight incubation is best.

Incubating the repair reaction at 37°C is recommended, however, a room temperature incubation will work nearly as well.

As with all PCR reactions, conditions may need to be optimized to achieve maximum amplicon yield.

Sequential Reaction Protocol:
This procedure is recommended if optimal PreCR DNA repair is desired and a PCR buffer other than ThermPol buffer is used for the PCR reaction.

The following protocol is recommended for a 50 µl PCR reaction:

1. At room temperature, combine 1X ThermPol Buffer, 100 µM dNTPs, 1X NAD+, 50–500 ng of the damaged template DNA and H2O to 49 µl.
2. Add 1 µl of the PreCR Repair Mix, and mix gently.
3. Incubate the repair reaction for 15–20 minutes at 37°C.
4. Place the reactions on ice.
5. The DNA repaired by the PreCR Repair Mix can now be used as a template for the subsequent PCR reaction in the desired buffer. Testing various amounts of the repaired template DNA in a 50 µl reaction is recommended, with 5 µl as a starting point.

Notes: It is common for PCR inhibitors to co-purify with DNA from degraded samples. One method to overcome this is to use BSA to bind the inhibitor and prevent it from interfering with PCR. If PCR inhibitors are suspected to be present in your samples we recommend that the supplied BSA be added to both the repair and PCR reactions to a final concentration of 1 mg/ml. Because the supplied BSA is 10 mg/ml this would require adding 5 µl BSA in a 50 µl reaction. If possible, it is best to perform both the repair and PCR reactions with and without BSA.

Control Reaction Protocol:
PCR conditions will vary depending on the PCR polymerase and PCR buffer. The control reaction was optimized for Taq DNA polymerase with ThermPol buffer (NEB#M0267). If using another polymerase or buffer it may be necessary to optimize the reaction conditions. The forward and reverse control PCR primers, the L1 Primer Mix, come pre-mixed at a concentration of 20 µM of each. It is important to run a negative control at the same time as the repair reaction. For the negative control add 1 µl of H2O instead of 1 µl of PreCR Repair Mix at the appropriate time. This is the unrepaired DNA reaction for comparison.

The following protocol is for a 50 µl PCR reaction.
1. At room temperature, combine 38 µl H2O, 5 µl ThermPol Buffer, 0.5 µl 100X NAD+, 0.5 µl of 10 mM dNTPs, and 3 µl of the supplied UV damaged Lambda DNA.
2. Add 1 µl PreCR Repair Mix and mix by gently pipetting up and down 3 times.
3. Incubate the repair reaction for 15 minutes at 37°C.
4. Place the reaction on ice.
5. Directly to the repair reaction, add 1 µl of the control primers, 0.5 µl of 10 mM dNTPs and 0.5 µl of Taq DNA polymerase (5 units/µl, not supplied). Mix gently.

6. Place into a thermocycler running the following program once the block temperature reaches > 90°C. Thermocycler program: 2 min at 95°C for 1 cycle, then 10 sec at 95°C, 30 sec at 65°C, 1 min at 72°C for 25 cycles and finally a 4°C hold. A PCR product of 1 kb should be seen.