

## Bst DNA Polymerase, Large Fragment



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M0275S 051150317031

# M0275S



**1,600 units**      **8,000 U/ml**      **Lot: 0511503**

**RECOMBINANT**    **Store at -20°C**    **Exp: 3/17**

**Description:** *Bst* DNA Polymerase, Large Fragment is the portion of the *Bacillus stearothermophilus* DNA Polymerase protein that contains the 5' → 3' polymerase activity, but lacks 5' → 3' exonuclease activity.

**Source:** *Bst* DNA Polymerase, Large Fragment is prepared from an *E. coli* strain containing a genetic fusion of the *Bacillus stearothermophilus* DNA Polymerase gene, lacking the 5' → 3' exonuclease domain, and the gene coding for

*E. coli* maltose binding protein (MBP). The fusion protein is purified to near homogeneity and the MBP portion of the fusion is cleaved off *in vitro*. The remaining polymerase is purified free of MBP (1).

### Applications:

- Isothermal amplification (LAMP)
- DNA sequencing through high GC regions (2,3)
- Rapid Sequencing from nanogram amounts of DNA template (4)

Supplied in: 50 mM KCl, 10 mM Tris-HCl (pH 7.5), 0.1 mM EDTA, 1 mM dithiothreitol, 0.1% Triton® X-100 and 50% glycerol.

### Reagents Supplied with Enzyme:

10X ThermoPol® Reaction Buffer.  
100 mM MgSO<sub>4</sub>

### 1X ThermoPol Reaction Buffer:

20 mM Tris-HCl  
10 mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>  
10 mM KCl  
2 mM MgSO<sub>4</sub>  
0.1% Triton X-100  
pH 8.8 @ 25°C

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**Unit Definition:** One unit is defined as the amount of enzyme that will incorporate 10 nmol of dNTP into acid insoluble material in 30 minutes at 65°C.

**Unit Assay Conditions:** 50 mM KCl, 20 mM Tris-HCl (pH 8.8), 10 mM MgCl<sub>2</sub>, 30 nM M13mp18 SS DNA, 70 nM M13 sequencing primer (-47) 24 mer, 200 μM dATP, 200 μM dCTP, 200 μM dGTP, 100 μM dTTP including [<sup>3</sup>H]-dTTP and 100 μg/ml BSA.

**Heat Inactivation:** 80°C for 20 minutes.

### Protocol:

#### Typical LAMP Protocol

Incubate the following reaction at 65°C for 30–60 minutes.

COMPONENT	25 μl REACTION	FINAL CONC.
10X ThermoPol Buffer	2.5 μl	1X (contains 2 mM MgSO <sub>4</sub> )
MgSO <sub>4</sub> (100 mM)	1.5 μl	6 mM (8 mM total)
dNTP Mix (10 mM)	3.5 μl	1.4 mM each
FIP/BIP Primers (25X)	1 μl	1.6 μM
F3/B3 Primers (25X)	1 μl	0.2 μM
LoopF/B Primers (25X)	1 μl	0.4 μM
<i>Bst</i> DNA Polymerase, Large Fragment (8,000 U/ml)	1 μl	320 U/ml
DNA Sample	variable	> 10 copies or more
Nuclease-free Water	to 25 μl	
Total Reaction Volume		25 μl

### General Guidelines:

1. A LAMP Primer Mix can be prepared with all 4 or 6 (with Loop) primers. A 25X Primer Mix should contain: 40 μM FIP, 40 μM BIP, 5 μM F3, 5 μM B3, 10 μM LoopF, 10 μM LoopB in TE or water.

(see other side)

CERTIFICATE OF ANALYSIS

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(see other side)

CERTIFICATE OF ANALYSIS

- Reactions should be setup on ice. If room temperature setup is desired, use *Bst* 2.0 WarmStart® DNA Polymerase (NEB #M0538).
- If analyzing via agarose gel electrophoresis or other method requiring opening LAMP reaction vessels, setup secondary analysis area and equipment to avoid contamination.
- Running a no-template control is strongly recommended to ensure amplification specificity.
- If optimization is desired, try titrating Mg<sup>2+</sup> (4–10 mM final) or *Bst* DNA Polymerase, Large Fragment (0.04–0.32 U/μl), or changing reaction temperature (50–68°C).

### Quality Control Assays

**16-Hour Incubation:** Incubation of a 50 μl reaction in ThermoPol Reaction Buffer containing a minimum of 500 units of *Bst* DNA Polymerase, Large Fragment with 1 μg of λ DNA for 16 hours at 65°C results in no detectable change in DNA banding pattern as determined by agarose gel electrophoresis.

Page 2 (M0275)

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Page 2 (M0275)

**Exonuclease Activity:** Incubation of a 50 μl reaction in ThermoPol Reaction Buffer containing a minimum of 500 units of *Bst* DNA Polymerase, Large Fragment with 1 μg of a mixture of single and double-stranded [<sup>3</sup>H] *E. coli* DNA for 4 hours at 65°C releases < 0.1% of the total radioactivity.

**Endonuclease Activity:** Incubation of a 50 μl reaction in ThermoPol Reaction Buffer containing a minimum of 500 units of *Bst* DNA Polymerase, Large Fragment with 1 μg of supercoiled φX174 DNA for 4 hours at 65°C results in < 10% conversion to the nicked form as determined by agarose gel electrophoresis.

**Physical Purity:** Purified to > 95% homogeneity as determined by SDS-PAGE analysis using Coomassie Blue detection.

### Enzyme Properties

#### Activity in NEBuffers

ThermoPol Buffer	125%
Unit Assay Conditions	100%
NEBuffer 1	50%
NEBuffer 2	100%
NEBuffer 3	50%
NEBuffer 4	100%

NEBuffers 1, 2, 3 and 4 must be supplemented with 0.1% Triton X-100 or 100 μg/ml BSA.

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Approximately 10% activity is observed in these buffers in the absence of BSA or Triton X-100.

**Notes On Use:** *Bst* DNA Polymerase does not exhibit 3'→5' exonuclease activity.

100 μg/ml BSA or 0.1% Triton X-100 is required for long term storage. Reaction temperatures above 70°C are not recommended.

*Bst* DNA Polymerase, Large Fragment cannot be used for thermal cycle sequencing or PCR.

### Companion Products Sold Separately:

Magnesium Sulfate (MgSO<sub>4</sub>) Solution  
#B1003S 6.0 ml

ThermoPol Reaction Buffer Pack  
#B9004S 6.0 ml

ThermoPol II (Mg-free) Reaction Buffer Pack  
#B9005S 6.0 ml

ThermoPol DF (Detergent-free) Reaction Buffer Pack  
#B9013S 6.0 ml

Deoxynucleotide Solution Set  
#N0446S 25 μmol each

Deoxynucleotide Solution Mix  
#N0447S 8 μmol each  
#N0447L 40 μmol each

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### References:

- Kong, H., Aliotta, J. and Pelletier, J.J., New England Biolabs, unpublished results.
- Griffin, H. and Griffin, A. (1994). *PCR Technology* (pp.228–229). Florida: CRC Press.
- McClary, J. et al. (1991) *J. DNA Sequencing and Mapping* 1, 173–180.
- Mead, D.A. et al. (1991) *BioTechniques* 11, 76–87.
- Notomi et al. (2000) *Nucleic Acids Res.* 28(12), E63.
- Hsieh et al. (2014) *Chem. Commun.* 50, 3747–3749.
- Tanner and Evans (2014) *Curr. Prot. Mol. Biol.* 105, 15.14.



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### References:

- Kong, H., Aliotta, J. and Pelletier, J.J., New England Biolabs, unpublished results.
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