The EpiMark Nucleosome Assembly Kit is used to make unmodified recombinant human nucleosomes with user-supplied DNA or the provided control DNA. Purified recombinant human Histone H2A/H2B Dimer and Histone H3.1/H4 Tetramer are mixed with DNA at 2 M NaCl. To generate nucleosomes, the salt concentration is lowered by dilution or dialysis allowing each histone tetramer to associate with two histone dimers and form the histone octamer on the DNA (1,2). A method for assaying nucleosome formation by gel shift assay is included. These nucleosomes may serve as a better substrate for enzymes that are inactive on the DNA or one of the core histones alone (3,4). Each reaction creates nucleosomes from ~1 µg of 208 bp DNA and may be scaled depending on the experiment.

Applications:
- ChiP assay
- HAT assay
- Enzymatic modification assay, (e.g., methylation)

Advantages:
- Highly pure, recombinant system
- Components stable for one year
- Pre-formed histone dimer and tetramer complexes simplify octamer formation
- Dilution protocol only requires a few hours for assembly

Kit Components:
- 20 µM Histone H2A/H2B Dimer 100 µl
- 10 µM Histone H3.1/H4 Tetramer 100 µl
- 10 µM Nucleosome Control DNA 20 µl

Quality Control
The EpiMark Nucleosome Assembly Kit has been tested using the control DNA provided as described in the Dilution Assembly Protocol. The control reaction yields a characteristic shift on 6% native polyacrylamide gel from 208 bp to ~700 bp.

Other Considerations
- Because stability can be an issue for the octamer in vitro without DNA, we provide the Dimer and Tetramer separately.
- Set up each reaction with dH₂O, 5 M NaCl, DNA and protein such that the final concentration is 2 M NaCl, noting that the Dimer and Tetramer are supplied in 2 M NaCl containing buffer.
- Always add the Dimer and Tetramer last!
- Because nucleosomes can dissociate when too dilute, it is recommended to keep the final protein concentration above 10 µg/ml (5).

Protocols
Two different protocols are available. The quickest is the dilution assembly protocol with assembly being ready in less than three hours. It is ideal for optimization since smaller volumes of all components may be used. The dialysis protocol does take longer but the concentration differences from starting material are minimized and it requires less sample handling.

Dilution Assembly Protocol

1. Set up each reaction with dH₂O, 5 M NaCl, DNA and protein such that the final concentration is 2.0 M NaCl (10 µM Histone H2A/H2B Dimer and Histone H3.1/H4 Tetramer).

General Guidelines

Molar Ratio
(2 Dimers : 1 Tetramer : 1 DNA Binding Site)

Because the correct ratio of DNA to protein determines the efficiency of an assembly, the kit components have been formulated to have equal volumes of each added in a reaction. After assembly, the reactions should contain 5–10% unbound DNA to mitigate aggregate formation which occurs when protein is in excess.

DNA Substrate
The Nucleosome Control DNA (208 bp) contains one possible binding site for an octamer. This enables visualization by gel shift assay due to the binding of the octamer and a characteristic shift from 208 bp to ~700 bp on a 6% native polyacrylamide gel. When using user-supplied substrate DNA, optimization may be required to determine the number of possible binding sites per molecule. A starting point for the amount of user supplied DNA to add per reaction can be determined using the formulas described in the DNA concentration formula section. Then, keeping the amount of DNA constant, the amount of octamer can be varied to find an optimal ratio of DNA to protein. To conserve DNA and protein during optimization, we would recommend the dilution protocol for 25 pmol.

For 50 pmol

This reaction, as described, can yield a maximum of 50 pmol nucleosome in 160 µl (0.3 pmol/µl; 0.3 µM nucleosome; 33.7 µg/ml protein).

1. Place 200 µl of Dilution buffer per reaction at room temperature.
2. Prepare the Reaction Assembly Mix on ice in the following order (for user-supplied substrate, suggested ratios have been included):

<table>
<thead>
<tr>
<th>0.5 to 1 Octamer to DNA</th>
<th>1 to 1 Octamer to DNA</th>
<th>1.5 to 1 Octamer to DNA</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control DNA only</td>
<td>0.5 µl</td>
<td>0.5 µl</td>
</tr>
<tr>
<td>10 µM Histone H2A/H2B Dimer</td>
<td>5.0 µl</td>
<td>5.0 µl</td>
</tr>
<tr>
<td>10 µM Histone H3.1/H4 Tetramer</td>
<td>5.0 µl</td>
<td>5.0 µl</td>
</tr>
<tr>
<td>2 M NaCl</td>
<td>0.5 µl</td>
<td>0.5 µl</td>
</tr>
<tr>
<td>5 M NaCl</td>
<td>0.5 µl</td>
<td>0.5 µl</td>
</tr>
<tr>
<td>10 µM Dimer</td>
<td>5.0 µl</td>
<td>5.0 µl</td>
</tr>
<tr>
<td>20 µM Tetramer</td>
<td>5.0 µl</td>
<td>5.0 µl</td>
</tr>
<tr>
<td>Total</td>
<td>20.0 µl</td>
<td>20.0 µl</td>
</tr>
</tbody>
</table>

The Dimer and Tetramer are supplied in 2 M NaCl.

3. Incubate reactions at room temperature for 30 minutes.
4. Add 7 µl room temperature dilution buffer to each reaction. This brings the reactions to 1.48 M NaCl, 27 µl total volume. Incubate at room temperature for 30 minutes.
5. Add 13 µl room temperature Dilution Buffer to each reaction. This brings the reactions to 1.0 M NaCl, 40 µl total volume. Incubate at room temperature for 30 minutes.
6. Add 27 µl room temperature Dilution Buffer to each reaction. This brings the reactions to 0.6 M NaCl, 67 µl total volume. Incubate at room temperature for 30 minutes.
7. Add 93 µl room temperature Dilution Buffer to each reaction. This brings the reactions to 0.25 M NaCl, 160 µl total volume. Incubate at room temperature for 30 minutes.
8. Store samples at 4°C.
9. Use gel shift assay to analyze samples.

(see other side)
Diaibol Assembly Protocol

This reaction can yield a maximum of 50 pmol nucleosome in 80 µl (0.3 pmol/µl; 0.3 µM nucleosome; 33.7 µg/ml protein).

1. Place 100 µl of Dilution Buffer per reaction at room temperature.
2. Prepare the Reaction Assembly Mix on ice in the following order (for user-supplied substrate, suggested ratios have been included):

<table>
<thead>
<tr>
<th>0.5 to 1</th>
<th>For 25 pmol DNA Substrate</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control DNA only (25 pmol)</td>
<td>0.5 to 1 to DNA</td>
</tr>
<tr>
<td>Water</td>
<td>0.5 µl</td>
</tr>
<tr>
<td>5M NaCl</td>
<td>2 µl</td>
</tr>
<tr>
<td>DNA</td>
<td>2.5 µl</td>
</tr>
<tr>
<td>20 µM Dimer</td>
<td>2.5 µl</td>
</tr>
<tr>
<td>10 µM Tetramer</td>
<td>2.5 µl</td>
</tr>
<tr>
<td>Total</td>
<td>10 µl</td>
</tr>
</tbody>
</table>

The Dimer and Tetramer are supplied in 2 M NaCl.

3. Incubate reactions at room temperature for 30 minutes.
4. Add 3.5 µl room temperature dilution buffer to each reaction. This brings the reactions to 1.48 M NaCl, 33.5 µg/ml protein. Incubate at room temperature for 30 minutes.
5. Add 6.5 µl room temperature dilution buffer to each reaction. This brings the reactions to 1.0 M NaCl, 20 µl total volume. Incubate at room temperature for 30 minutes.
6. Add 13.5 µl room temperature dilution buffer to each reaction. This brings the reactions to 0.6 M NaCl, 33.5 µl total volume. Incubate at room temperature for 30 minutes.
7. Add 46.5 µl room temperature dilution buffer to each reaction. This brings the reactions to 0.25 M NaCl, 80 µl total volume. Incubate at room temperature for 30 minutes.
8. Store samples at 4°C.

9. Use gel shift assay to analyze samples.

Gel Shift Assay

1. Mix 10 µl of each sample with 2 µl of 100% glycerol, including control reactions.
2. Load 10 µl of each sample onto a 6% DNA retardation gel. We recommend running 2 µl TriDye 100 bp DNA Ladder (NEB #N3271) alongside reactions.
3. Electrophorese according to gel manufacturer protocol in buffer containing no ethidium bromide. Using the marker lane to track the gel run, run the blue dye off the gel and the green dye about two thirds down the gel. Xylene Cyanol in these gels runs approximately where the 200 bp substrate runs.
4. Analyze gel.
5. Analyze gel.

Diaibol Concentration Formulas

To determine the concentration of binding sites:
1. 1.0 A260 unit double stranded DNA = 50 µg/ml
2. MW of double stranded DNA = # base pairs x 650 daltons/base pairs
3. pmol/µl of double stranded DNA = µg/µl DNA x 10^6 pg per µg/MW
4. µM double stranded DNA = pmol/µl of double stranded DNA
5. Size of DNA in bp / 150 bp = µM of potential binding sites
6. µM of binding sites = # potential binding sites x µM DNA
7. Place dialysis units in 1.5 M NaCl buffer for 2–3 hours at 4°C and then transfer to each consecutively lower NaCl concentration buffer for 2–3 hours at 4°C with either the 0.6 M or 0.25 M NaCl buffer dialysis being an overnight step.
8. Transfer the samples to tubes. The volume will have increased because of the salt dialysis. Equalize sample volumes to 150 µl using the 0.25 M NaCl buffer. If volumes are off by more than 20%, some thing may have gone wrong with the set up of those samples.
9. Store samples at 4°C.

References


Components Sold Separately:

Histone H2A/H2B Dimer Human, Recombinant #M2508S 1 nmol
Histone H3.1/H4 Tetramer Human, Recombinant #M2509S 1 nmol
Nucleosome Control DNA #N1202S 10 µM

NEW ENGLAND BIOLABS® and EPIMARK® are registered trademarks of New England Biolabs, Inc.
TRIDYE™ is a trademark of New England Biolabs, Inc.
This product is intended for research purposes only. This product is not intended to be used for therapeutic or diagnostic purposes in humans or animals.