NEB 5-alpha Competent E. coli (High Efficiency)

C2987H

20 x 0.05 ml/tube Lot: 221
Store at –80°C

CAUTION: This product contains DMSO, a hazardous material. Review the SDS before handling.

Description: Chemically competent E. coli cells suitable for high efficiency transformation in a wide variety of applications.

Features:
- DH5α™ derivative
- Transformation efficiency: 1–3 x 10⁹ cfu/µg pUC19 DNA
- Efficient transformation of unmethylated DNA derived from PCR, cDNA and many other sources (hisD78)
- Activity of nonspecific endonuclease 1 (endA1) eliminated for highest quality plasmid preparations
- Resistance to phage T1 (fhuA2)
- Suitable for blue/white screening by α-complementation of the β-galactosidase gene
- Reduced recombination of cloned DNA (recA1)
- K12 Strain
- Free of animal products

Reagents Supplied:
20 x 0.05 ml/tube of chemically competent NEB 5-alpha Competent E. coli cells (Store at –80°C)
20 ml of SOC Outgrowth Medium (Store at room temperature)
0.025 ml of 50 pg/µl pUC19 Control DNA (Store at –20°C)

Quality Control Assays
Transformation Efficiency: 100 pg of pUC19 plasmid DNA was used to transform NEB 5-alpha Competent E. coli following the high efficiency protocol provided. 1–3 x 10⁹ colonies formed/µg after an overnight incubation on LB-ampicillin plates at 37°C.
Untransformed cells were also tested for resistance to phage φ80, a standard test for resistance to phage T1 and sensitivity to ampicillin, chloramphenicol, kanamycin, nitrofurantoin, spectinomycin, streptomycin and tetracycline. The cells were shown to be suitable for blue/white screening by α-complementation of the β-galactosidase gene using pUC19.

High Efficiency Transformation Protocol
Perform steps 1–7 in the tube provided.
1. Thaw a tube of NEB 5-alpha Competent E. coli cells on ice for 10 minutes.
2. Add 1–5 µl containing 1 pg–100 ng of plasmid DNA to the cell mixture. Carefully flick the tube 4–5 times to mix cells and DNA. Do not vortex.
3. Place the mixture on ice for 30 minutes. Do not mix.
5. Place on ice for 2 minutes. Do not mix.
6. Pipette 950 µl of room temperature SOC into the mixture.
7. Place at 37°C for 60 minutes. Shake vigorously (250 rpm) or rotate.
8. Warm selection plates to 37°C.
9. Mix the cells thoroughly by flicking the tube and inverting, then perform several 10-fold serial dilutions in SOC.
10. Spread 50–100 µl of each dilution onto a selection plate and incubate overnight at 37°C. Alternatively, incubate at 30°C for 24–36 hours or 25°C for 48 hours.

5 Minute Transformation Protocol
The following protocol results in only 10% efficiency compared to the High Efficiency Transformation Protocol. Perform steps 1–6 in the tube provided.
1. Remove cells from –80°C freezer and thaw in your hand.
2. Add 1–5 µl containing 1 pg–100 ng of plasmid DNA to the cell mixture. Carefully flick the tube 4–5 times to mix cells and DNA. Do not vortex.
3. Place the mixture on ice for 2 minutes. Do not mix.
5. Place on ice for 2 minutes. Do not mix.
6. Pipette 950 µl of room temperature SOC into the mixture. Immediately spread 50–100 µl onto a selection plate and incubate overnight at 37–42°C. NOTE: Selection using antibiotics other than ampicillin may require some outgrowth before plating on selective media. Colonies develop faster at temperatures above 37°C, however some constructs may be unstable at elevated temperatures.

Transformation Protocol Variables
Thawing: Cells are best thawed on ice and DNA added as soon as the last bit of ice in the tube disappears. Cells can also be thawed by hand, but warming above 0°C will decrease the transformation efficiency.

Incubation of DNA with Cells on Ice: For maximum transformation efficiency, cells and DNA should be incubated together on ice for 30 minutes. Expect a 2-fold loss in transformation efficiency for every 10 minutes this step is shortened.

Heat Shock: Both the temperature and the timing of the heat shock step are important and specific to the transformation volume and vessel. Using the transformation tube provided, 30 seconds at 42°C is optimal.

Outgrowth: Outgrowth at 37°C for 1 hour is best for cell recovery and for expression of antibiotic resistance. Expect a 2-fold loss in transformation efficiency for every 15 minutes this step is shortened. SOC gives 2-fold higher transformation efficiency than LB medium; and incubation with shaking or rotating the tube gives 2-fold higher transformation efficiency than incubation without shaking.

Plating: Selection plates can be used warm or cold, wet or dry without significantly affecting the transformation efficiency. However, warm, dry plates are easier to spread and allow for the most rapid colony formation.

DNA Effects on Transformation Efficiency and Colony Output: The optimal amount of DNA to use in a transformation reaction is lower than commonly recognized. Using clean, supercoiled pUC19, the efficiency of transformation is highest in the 10 pg–1 ng range. However, the total colonies which can be obtained from a single transformation reaction increase up to about 100 ng.
Antibiotics for Plasmid Selection

**Omit IPTG for potentially toxic genes**

<table>
<thead>
<tr>
<th>Antibiotic</th>
<th>Working Concentration</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ampicillin</td>
<td>100 µg/ml</td>
</tr>
<tr>
<td>Carbenicillin</td>
<td>100 µg/ml</td>
</tr>
<tr>
<td>Chloramphenicol</td>
<td>33 µg/ml</td>
</tr>
<tr>
<td>Kanamycin</td>
<td>30 µg/ml</td>
</tr>
<tr>
<td>Streptomycin</td>
<td>25 µg/ml</td>
</tr>
<tr>
<td>Tetracycline</td>
<td>15 µg/ml</td>
</tr>
</tbody>
</table>

**Solutions/Recipes**

**SOB:**
2% Vegetable peptone (or Tryptone)
0.5% Yeast Extract
10 mM NaCl
2.5 mM KCl
10 mM MgCl₂
10 mM MgSO₄

**SOC:**
SOB + 20 mM Glucose

**LB agar:**
1% Tryptone
0.5% Yeast Extract
0.17 M NaCl
1.5% Agar

**Blue/White Screening:**
X-gal 80 µg/ml
IPTG* 0.3 mM

*Omit IPTG for potentially toxic genes

**DNA Contaminants to Avoid**

- Detergents
- Phenol
- Ethanol or Isopropanol
- PEG*
- DNA binding proteins*

*Ideally, DNA for transformation should be purified and resuspended in water or TE. However, up to 10 µl of DNA directly from a ligation mix can be used with only a two-fold loss of transformation efficiency. Where it is necessary to maximize the number of transformants (e.g. a library), a purification step, either a spin column or phenol/chloroform extraction and ethanol precipitation should be added.

**Calculating Transformation Efficiency**

Transformation efficiency is defined as the number of colony forming units (cfu) which would be produced by transforming 1 µg of plasmid into a given volume of competent cells. The term is somewhat misleading in that 1 µg of plasmid is rarely actually transformed. Instead efficiency is routinely calculated by transforming 100 pg–1 ng of highly purified supercoiled plasmid under ideal conditions. If you plan to calculate efficiency to compare cells or ligations, keep in mind the many variables which affect this metric.

Transformation efficiency (TE) equation:

\[ TE = \frac{\text{Colonies}}{\mu g \times \text{Dilution}} \]

Colonies = the number of colonies counted on the plate
µg = the amount of DNA transformed expressed in µg
Dilution = the total dilution of the DNA before plating

TE calculation example:

Transform 2 µl (100 pg) of control pUC19 DNA into 50 µl of cells, outgrow by adding 250 µl of SOC and dilute 10 µl up to 1 ml in SOC before plating 30 µl. If you count 150 colonies on the plate, the TE is:

\[ \text{Colonies} = 150 \]
\[ \mu g \text{ DNA} = 0.0001 \]
\[ \text{Dilution} = 10/3000 \times 30/1000 = 0.001 \]
\[ TE = 150/0.0001/0.001 = 1.5 \times 10^9 \text{ cfu/µg} \]

**Strain Properties**

The properties of this strain that contribute to its usefulness as a cloning strain are described below. The genotypes underlying these properties appear in parentheses.

Blue/White Screening (ω80ΔargF-lacZ)M15: makes the ω-fragment of β-galactosidase (β-gal); (argF-lacZ) deletes the β-gal gene on the chromosome, pUC19 and similar plasmids code for the α-peptide of β-gal (lacZ).

The α-peptide can combine with the ω-fragment of β-gal that is carried on ω80 (α-complementation). When β-gal is reconstituted in this manner it can cleave 5-bromo-4-chloro-3-indolyl-β-D-galactosidase (X-gal) and results in blue colonies on an X-gal plate. Inserts cloned into the plasmid polylinker disrupt the α-peptide gene and the colonies are white.

Recombination Deficient (recA1): E. coli has a repair system that will recombine homologous sequences. Genomic clones often have duplicated regions, and RecA mediated rearrangements can be problematic, particularly when regions of homology are longer than 50 bp. Strains which have the RecA function deleted tend to grow more slowly than recA- strains.

Endonuclease I Deficient (endA1): The periplasmic space of wild type E. coli cells contains a nonspecific endonuclease. Extreme care must be taken to avoid degradation of plasmids prepared from these cells. The endA mutation deletes this endonuclease and can significantly improve the quality of plasmid preparations.

Restriction Deficient (hsdR17): Wild type E. coli K12 strains carry a restriction endonuclease which cleaves DNA with sites (AAC(N6)GTGC and GCAC(N6)GTT). While E. coli DNA is protected from degradation by a cognate methyl-transferase, foreign DNA will be cut at these sites. The hsdR mutation eliminates this endonuclease activity. However, this strain has functional methyl restriction systems and may not be suitable for direct cloning of eukaryotic DNA.

T1 Phage Resistant (thuA2): T1, an extremely virulent phage requires the E. coli ferric hydroxamate uptake receptor for infectivity. Deletion of this gene confers resistance to this type of phage, but does not significantly affect the transformation or growth characteristics of the cell.

---

DH5α™ is a trademark of Invitrogen Corporation.

**Companion Products Sold Separately:**

SOF Outgrowth Medium #B9020S 4 x 25 ml medium

---

**Genotype:** thuA2 Δ(argF-lacZ)U169 phoA glnV44 ω80Δ[argF-lacZ]M15 gyrA96 recA1 relA1 endA1 thi-1 hsdR17